SPIROKETALS: THE SYNTHESIS OF AN OLIVE FLY PHEROMONE COMPONENT, 4-HYDROXY-1,7-DIOXASPIRO[5,5]UNDECANE, VIA A NOVEL CATION-OLEFIN CYCLISATION STEP

I. Trevor Kay \star and Emyr G. Williams

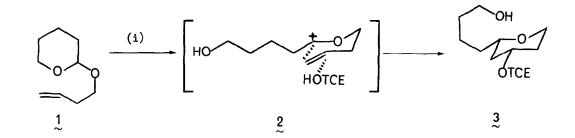
ICI Plant Protection Division, Jealott's Hill, Bracknell, Berkshire RG12 6EY

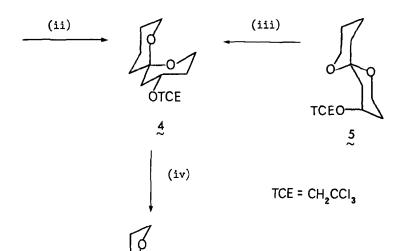
<u>Summary</u>: An acid-catalysed rearrangement of the THP-ether of homoallylic alcohol gives ready access to an 0-protected derivative of 4-hydroxytetrahydropyran and thence by two further steps to provide a short and highly stereoselective route to the title spiroketal.

The synthesis of spiroketals, particularly those containing hydroxyl groups has been the subject of much recent work.¹⁻⁹ The interest in these spirocycles has been stimulated by their occurrence as structural elements of several important antibiotics¹⁰ as well as their identification as insect pheromones.¹¹ We have developed a new and highly stereoselective route to 4-hydroxy-1,7-dioxaspiro[5,5]undecanes and report here its application to the synthesis of a pheromone component¹ <u>6</u> of the female olive fly Dacus cleae.

Hitherto, most¹⁻⁶ approaches to these spiroketals have involved as the key step the addition of carbanions to δ -valerolactones followed by cyclisation of the resultant lactols. Our route (SCHEME) is fundamentally different; the enabling transformation 1-[2] -3 utilises a cation-olefin cyclisation as in 2 to provide a stereoselective route to the protected 4-hydroxytetrahydropyran 3. To our knowledge such a transformation of a THP-ether of an homoallylic alcohol with cleavage of one tetrahydropyran ring and the formation of another has not been reported previously. Thus a mixture of the ether 1 (1 mol), trichloroethanol (3.5 mol) and BF₃OEt₂ (0.5 mol) when kept at room temperature for 18 hr. gave after work up and short-path distillation a 61% yield of the oily product 3¹² as a single¹³ isomer. An advantage of using trichloroethanol (rather than, say, formic acid which gives lack of regiospecificity arising from transesterification) as the cation-trapping reagent is that it provides a secure, but easily removeable, 0-protecting group leaving the primary -OH group of 3 exposed for further elaboration.

The spirocyclisation of 3 was accomplished via its hypoiodite 14 by heating (ca. 30 min) the compound under reflux in cyclohexane together with iodine (1 equivalent) and mercuric oxide. Under these non-acidic conditions there was obtained mainly the desired spiroketal 45 together with variable (up to 30%) amounts of its isomer 5. Brief exposure of the mixture (in cyclohexane) to a catalytic quantity of TFA brought about the complete (C-6) isomerisation of 5 to the less-hindered 4. The formation of 5 probably reflects the cationic (planar) nature of C-6 prior to spirocyclisation. The overall conversion of 3.44 was 50%.





́бн ~€



<u>Reagents</u>: (i) HOTCE, BF₃OEt₂; (ii) I₂, HgO; (iii) TFA; (iv) Zn-HCO₂H

Clean removal of the TCE protecting group from $\frac{4}{2}$ proved at first to be unexpectedly difficult due to the intervention of the difficultly reduced dichloroethoxy¹⁶ derivative formed during the zinc-acetic acid reduction. The use of zinc-formic acid overcame this problem. Thus brief (ca. 15 min.) stirring of $\frac{4}{2}$ in 9:1 formic acid and water (NaOAc) together with an excess of activated¹⁷ zinc dust gave (86%) the racemic pheromone component $\underline{6}$ containing less than 2% of its (C-6) isomer¹⁸ ($\underline{5}$; TCE=H).

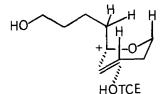
The ¹H-NMR and glc retention times for <u>6</u> were as reported ¹ for the natural product.

We thank Professor R. Baker for an authentic sample of $\underline{6}$ provided for comparative purposes.

References and notes

- 1. R. Baker, R.H. Herbert, and A.H.Parton, J.C.S.Chem.Comm., 601 (1982).
- 2. R. Baker, R.H.O.Boyes, D.M.P.Broom, J.A. Devlin, and C.J.Swain, *ibid*, 829 (1983).
- 3. S.V.Attwood, A.G.M.Barrett, and J.-C.Floret, ibid, 556 (1982).
- A.B.Smith, S.R.Schow, L.D.Bloom, A.S.Thompson, and K.N.Winzenberg, <u>J.Am.Chem.Soc.</u>, <u>104</u>, 4015 (1982).
- 5. D.R.Williams, B.A.Barner, K. Nishitani, and J.G.Phillips, *ibid*, 104, 4708 (1982).
- 6. D.R.Williams and B.A.Barner, Tetrahedron Lett., 427 (1983).
- 7. P. Kocienski and C. Yeates, *ibid*, 3905 (1983).
- 8. S.V.Ley and B.Lygo, ibid, 4625 (1982).
- 9. R.E.Ireland and J.P.Daub, J.Org.Chem., <u>48</u>, 1303 (1983).
- For example, milbemycins: H.Mishima, M.Kurabayashi, C.Tamura, S.Sato, H.Kuwano and A.Saito, <u>Tetrahedron Lett.</u>, 711 (1975); avermectins: G.Albers-Schonberg, B.H.Arison, J.C.Chabala, A.W.Douglas, P.Eskola, M.H.Fisher, A.Lusi, H.Mrozik, J.L.Smith, and R.L.Tolman, <u>J.Am.Chem.Soc.</u>, <u>103</u>, 4216 (1981); monensin: G.Agtarap, J.W.Chamberlain, M. Pinkerton, and L. Steinrauf, <u>ibid</u>, <u>89</u>, 5737 (1967); talaromycins: D.G.Lynn, N.J.Phillips, W.C.Hutton, J.Shabanowitz, D.I.Fennell, and R.J.Cole, <u>ibid</u>, <u>104</u>, 7319 (1982).
- W.Francke, W.Reith, G.Bergstrom, and J.Tengo, <u>Naturwis.</u>, <u>67</u>, 149 (1980); R.Baker, R.Herbert, P.E.Howse, O.T.Jones, W.Francke, and W.Reith, <u>J.C.S.Chem.Comm.</u>, 52 (1980).
- 12. ^LH-NMR, (CDCl₃, 90 MHz); 4.12(2H,s),4.0(1H,ddd,J=11.7,5.0,2.5 Hz),3.8(1H,tt,J=10, 5 Hz),3.64(2H,t,J=7 Hz),3.40(1H,td,J=11.7,2.5 Hz),3.24(1H,m),2.1-1.1(11H,m).

13. The alternative mode of cyclisation (below) is presumably disfavoured on steric grounds:



A similar argument has been advanced to account for the stereoselectivity observed in a perhydrohistrionicotoxin synthesis: H.E.Schoemaker and W.N.Speckamp, <u>Tetrahedron</u>, <u>36</u>, 951 (1980).

- 14. M.L.Mihailovic, S.Gojkovic, and S. Konstantinovic, <u>ibid</u>, <u>29</u>, 3675 (1973).
- 15. ¹H-NMR, (CDCl₃, 90 MHz): 4.08(2H,s),4.04(1H,tt,J=10,5 Hz),3.8-3.5(4H,m),2.28-1.30 (10H,m).
- The reduction of trichloroethoxy- to their dichloroethoxy-derivatives has previously been noted but only for electrochemical reduction: M.F.Semmelhack and G.E.Heinsohn, J.Am.Chem.Soc., 94, 5139 (1972).
- 17. Just prior to use the zinc dust was stirred for 10 min with dilute HCl, washed with water then acetone and dried <u>in vacuo</u>.
- 18. This is probably formed as the result of the exposure of 5 to the acidic medium (cf. reference 1.) during removal of the protecting group.

(Received in UK 19 October 1983)